Fenebrutinib, a Noncovalent, Reversible, Bruton's Tyrosine Kinase Inhibitor, Potently Blocks Neuroinflammation Induced by FcyR Activation in Human Microglial Systems: Implications for MS Treatment

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OBJECTIVE

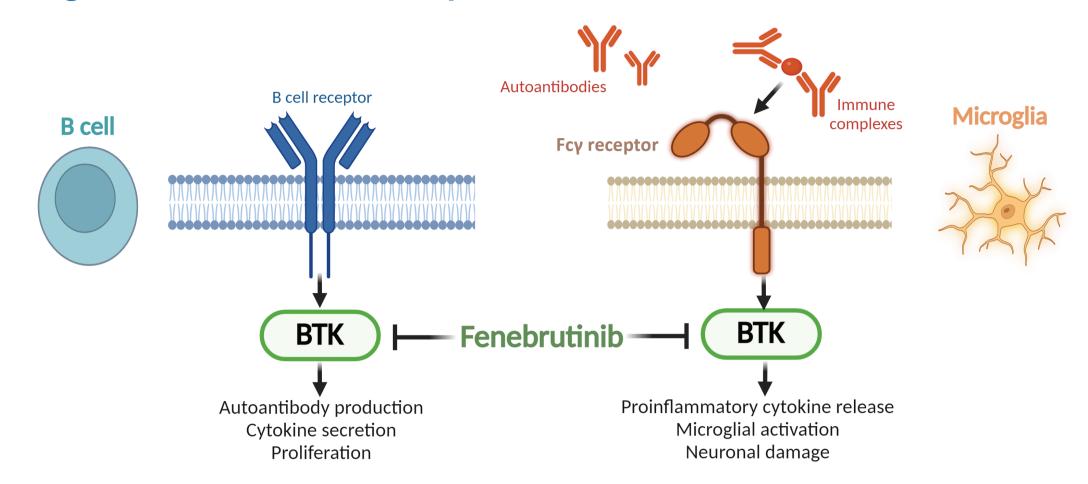
To evaluate the pharmacological and functional properties of the BTKi FEN in human microglial systems.

CONCLUSIONS

INTRODUCTION

- Neuroinflammation driven by detrimental microglial activation may contribute to progressive disease biology in **multiple sclerosis** (MS)
- In the periphery, **Bruton's tyrosine kinase** (BTK) is expressed in both adaptive (B cells) and innate (monocytes, macrophages) immune cells,¹ whereas in

Figure 1. Fenebrutinib Proposed Mechanism of Action



FEN acts on microglia to ameliorate pathogenic neuroinflammation resulting from FcyR activation. In people with MS, this effect may reduce progressive neurodegeneration resulting from chronic CNS inflammation in MS.

Further biomarker and efficacy assessments in clinical trials may help to confirm the clinical relevance of this observed effect on microglia.

the central nervous system (CNS), BTK is expressed primarily in **microglia**²

- Fenebrutinib (FEN) is a potent, highly selective, noncovalent, reversible BTK inhibitor (BTKi)³ that may address MS disease progression by targeting multiple pathogenic mechanisms in both the peripheral and CNS arms of the immune system
- Much research to date has investigated the effects of BTKis using rodent disease models. Here, we characterise the impact of FEN in blocking BTK-mediated, disease-associated Fcy receptor (FcyR) activation in human microglia and complex human brain cell systems to enhance translatability to people with MS

RESULTS

Figure 2. FEN Potently Blocked FcyR-Induced Activation in Human Microglia

• FEN showed similar high potency in microglia to B cells and monocytes

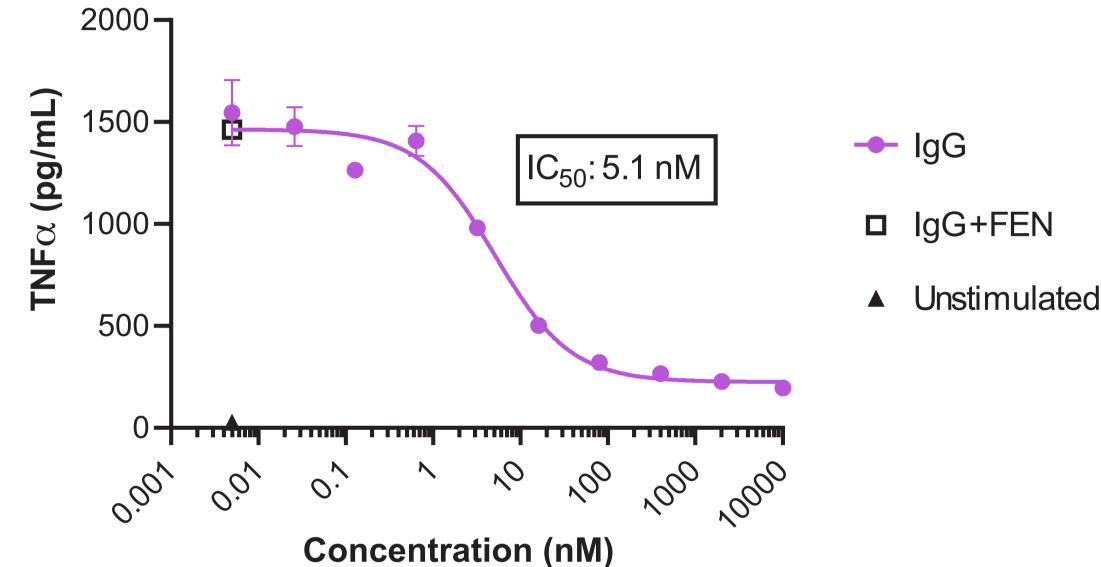
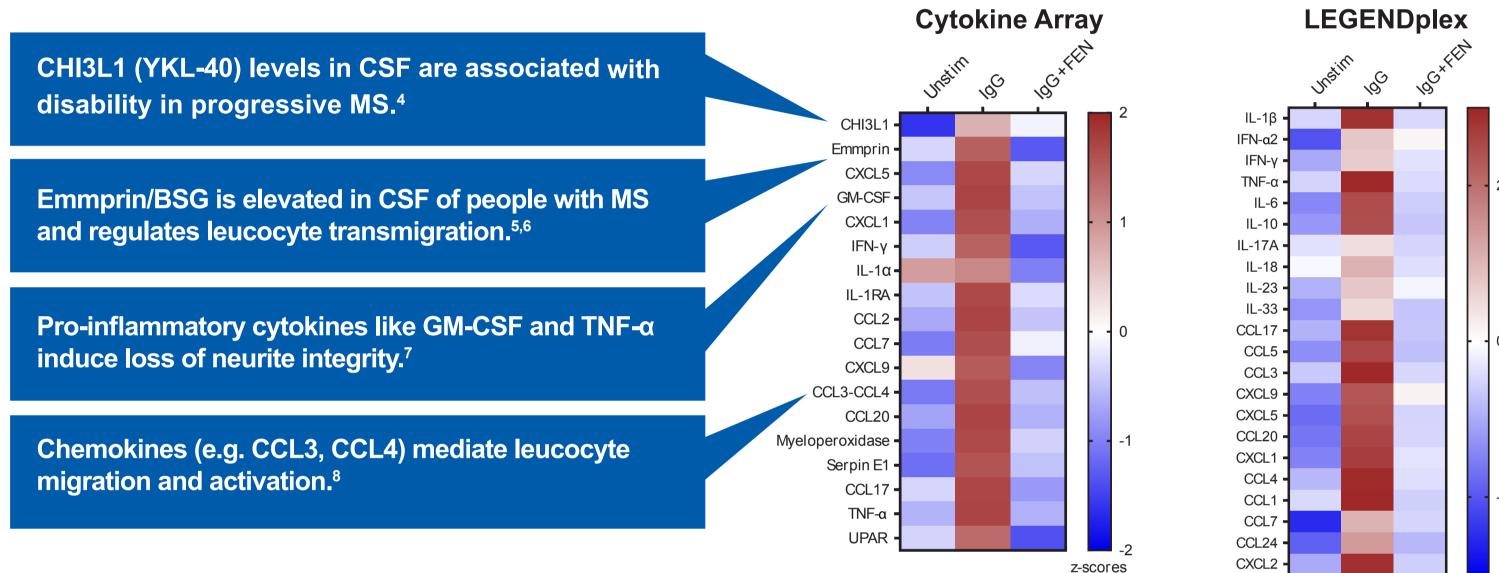


Figure 3. FEN Acts on Human Microglia to Attenuate the Release of Multiple Pro-Inflammatory **Cytokines and Chemokines Linked to MS Pathogenesis**



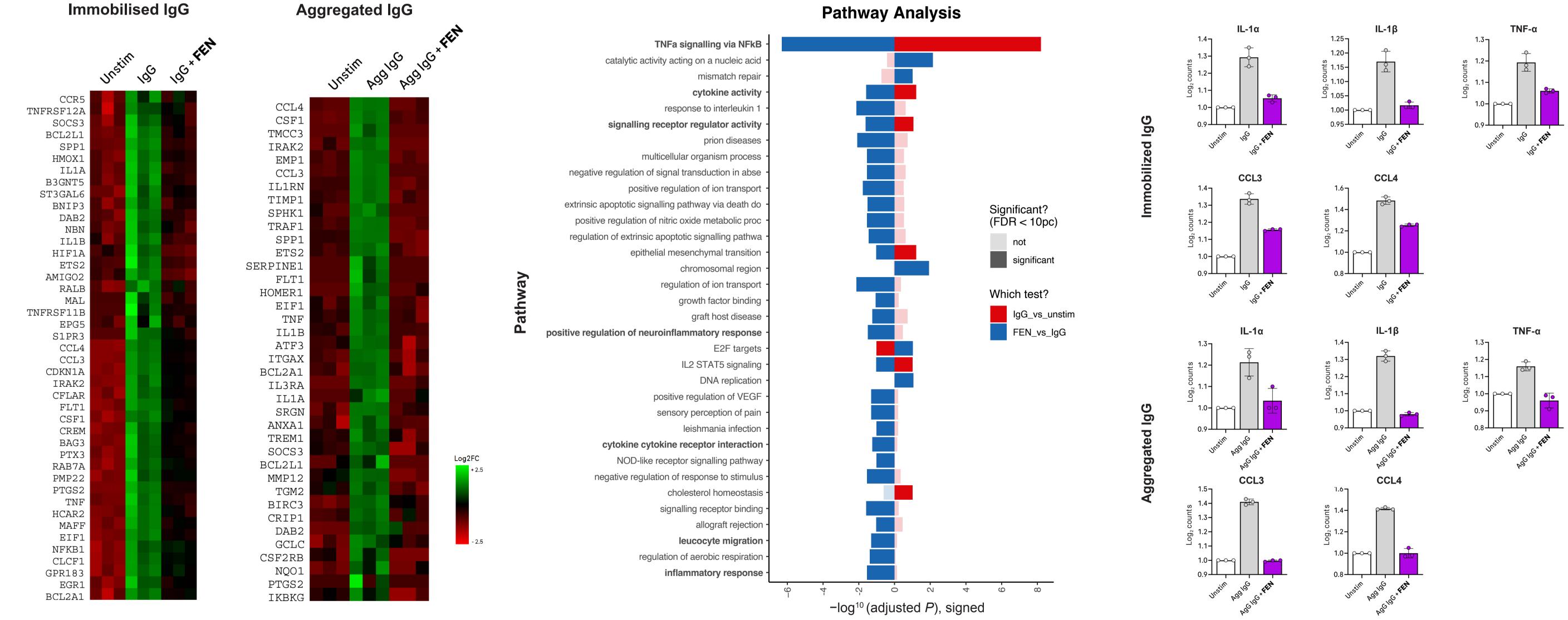
BSG, basigin; CCL, C-C motif chemokine ligand; CHI3L1, chitinase 3 like 1; CSF, cerebrospinal fluid; FEN, fenebrutinib; CXCL, C-X-C chemokine ligand; GM-CSF, granulocyte macrophage colony-stimulating factor; IFN, interferon; IgG, immunoglobulin G; IL, interleukin; MS, multiple sclerosis; TNF-α, tumour necrosis factor α; UPAR, urokinase plasminogen activator receptor.

Figure 4: Gene Expression Analyses Identified Microglial Genes and Pathways Linked to Neuroinflammation and Cytokine Signalling Modulated by FEN

FEN reversed gene expression changes induced by FcγR signalling in microglia

• Consistent effects of FEN on cytokine (IL-1α, IL-1β, TNF-α) and chemokine (CCL3, CCL4) genes were observed

• Transcriptional signatures of FEN activity included pathways linked to neuroinflammation and cytokine signalling

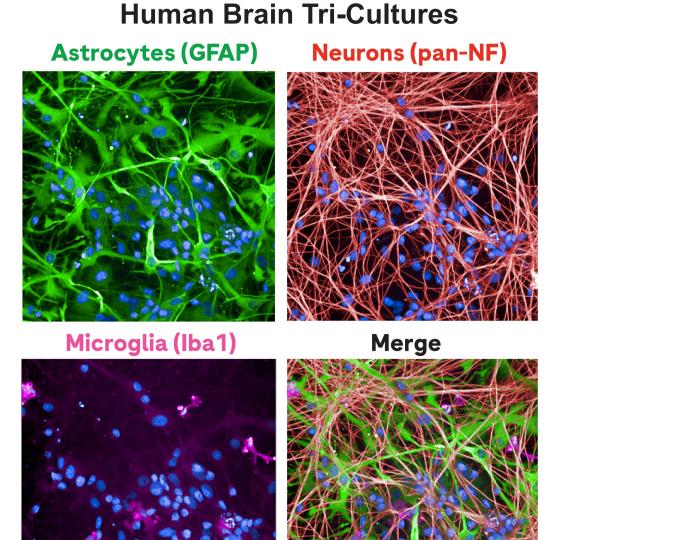


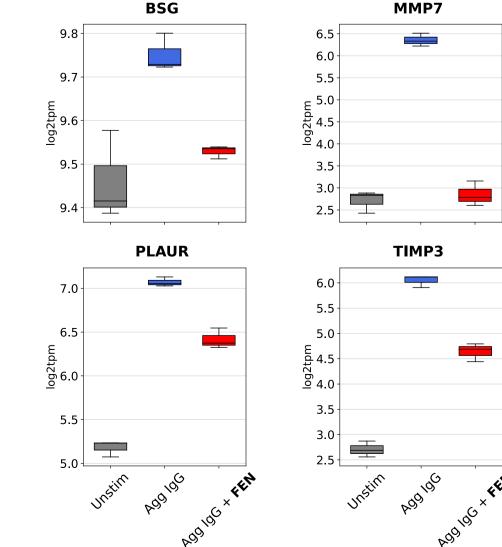
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Agg, aggregated; FDR, false discovery rate; FEN, fenebrutinib; IgG, immunoglobulin G.

Figure 5. FEN Blocks FcyR-Mediated Activation of Basigin and Matrix Metalloproteinase Pathways in Human Brain Tri-Culture System

- BSG (an MMP inducer) and MMP are elevated in MS plaques, and BSG blockade attenuates the severity of experimental autoimmune encephalomyelitis, an animal model of MS⁹
- BBB damage by MMPs mediates transmigration of leukocytes into CNS¹⁰





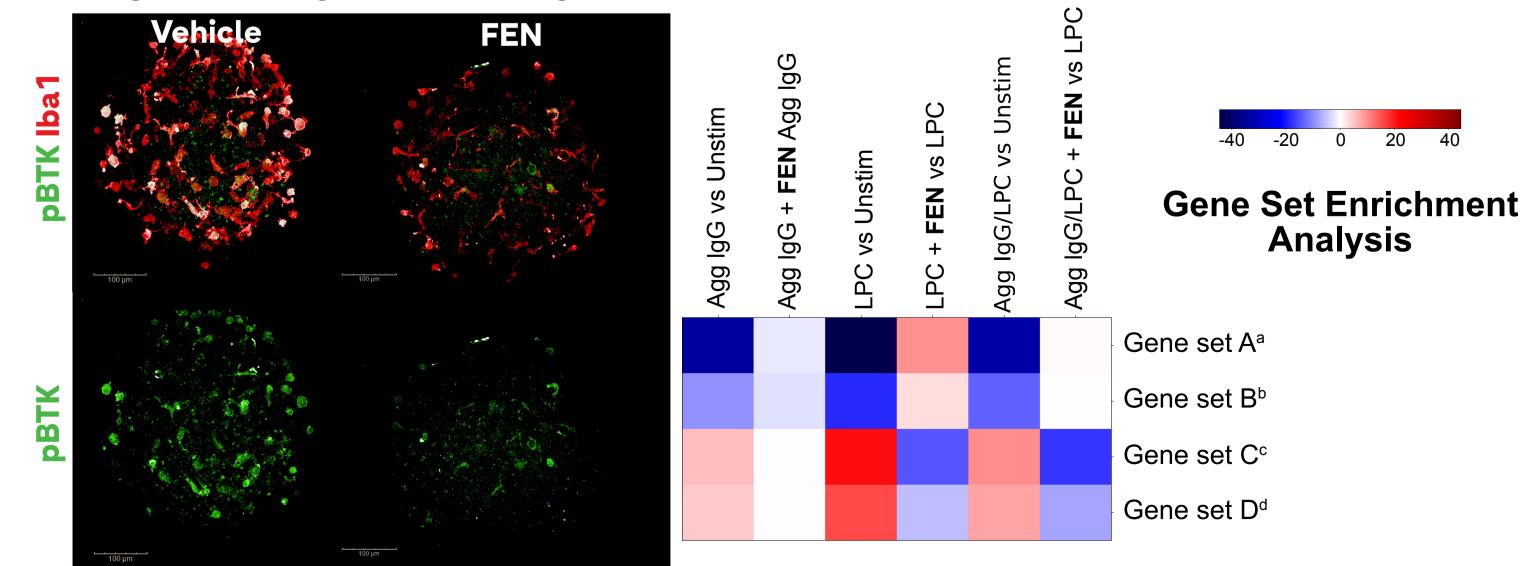
Targets of BSG

Agg, aggregated; BSG, basigin; GFAP, glial fibrillary acidic protein; Iba1, ionized calcium binding adaptor molecule 1; IgG, immunoglobulin G; MMP, matrix metalloproteinase; NF, neurofilament; PLAUR, plasminogen activator, urokinase receptor; TIMP3, TIMP metallopeptidase inhibitor 3.

Figure 6. Treatment of Immunocompetent Human Brain Organoids With FEN Blocked **FcyR-Mediated Inflammatory Signalling**

• FEN reversed FcyR- and lysolecithin (LPC)-mediated effects on cholesterol metabolism and toll-like receptor (TLR) inflammatory signalling pathways

Microglia-Containing Human Brain Organoids



Agg, aggregated; BTK, Bruton's tyrosine kinase; FEN, fenebrutinib; Iba1, ionized calcium binding adaptor molecule 1; IgG, immunoglobulin G. ^aGene set A = cholesterol_biosynthesis_REACTOME. ^bGene set B = activation_of_gene_expression_by_SREBF_SREBP_REACTOME. ^cGene set C = enriched_in_monocytes_(II)_(M11.0). ^dGene set D = TLR_and_inflammatory_signalling_(M16).

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DISCLOSURES

J Langlois, S Lange, J DeGeer, W Macnair, M Ebeling, R Schmucki, L Collin and J Keaney are employees and shareholders of F. Hoffmann-La Roche Ltd. C Harp and Y-A Shen are employees of Genentech, Inc. and shareholders of F. Hoffmann-La Roche Ltd. JA Nicholas has received research grants from Biogen, Novartis, PCORI, Genentech, Inc., University of Buffalo; she has received compensation for consulting from EMD Serono, Genentech, Inc., Greenwich Biosciences, Novartis, TG Therapeutics and Sanofi and speaking honoraria from BMS, EMD Serono, Horizon, TG Therapeutics. **J Oh** has received personal compensation for serving as a consultan from Biogen, Bristol-Myers Squibb, Eli Lilly, Novartis AG, EMD Serono, F. Hoffmann-La Roche Ltd and Sanofi. The institution of Dr Oh has received research support from Biogen, EMD Serono and F. Hoffmann-La Roche Ltd.

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Supplementary Material

Methods



- Efficacy and potency of fenebrutinib (FEN) was assessed in human-induced pluripotent stem cell (iPSC)–derived microglia by measuring tumour necrosis factor α (TNF-α) release following Fcγ receptor (FcγR) activation
- Two different immunoassay methods were used to detect a broad range of cytokines and chemokines modulated by FEN treatment
- Two different methods to activate FcγR signaling in human microglia were used: immobilized immunoglobulin G (IgG) and aggregated IgG
- NanoString gene expression profiling and RNA-sequencing was used to capture transcriptional signatures associated with FEN treatment in iPSC-derived microglia, human brain tri-cultures and human brain organoids